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# Actin pedestal formation by enteropathogenic *Escherichia coli* and intracellular motility of *Shigella flexneri* are abolished in N-WASP-defective cells

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In mammalian cells, actin dynamics is tightly controlled through small GTPases of the Rho family, WASP/Scar proteins and the Arp2/3 complex. We employed Cre/*loxP*-mediated gene targeting to disrupt the ubiquitously expressed N-WASP in the mouse germline, which led to embryonic lethality. To elucidate the role of N-WASP at the cellular level, we immortalized embryonic fibroblasts and selected various N-WASP-defective cell lines. These fibroblasts showed no apparent morphological alterations and were highly responsive to the induction of filopodia, but failed to support the motility of *Shigella flexneri*. In addition, enteropathogenic *Escherichia coli* were incapable of inducing the formation of actin pedestals in N-WASP-defective cells. Our results prove the essential role of this protein for actin cytoskeletal changes induced by these bacterial pathogens *in vivo* and in addition show for the first time that N-WASP is dispensible for filopodia formation.

## INTRODUCTION

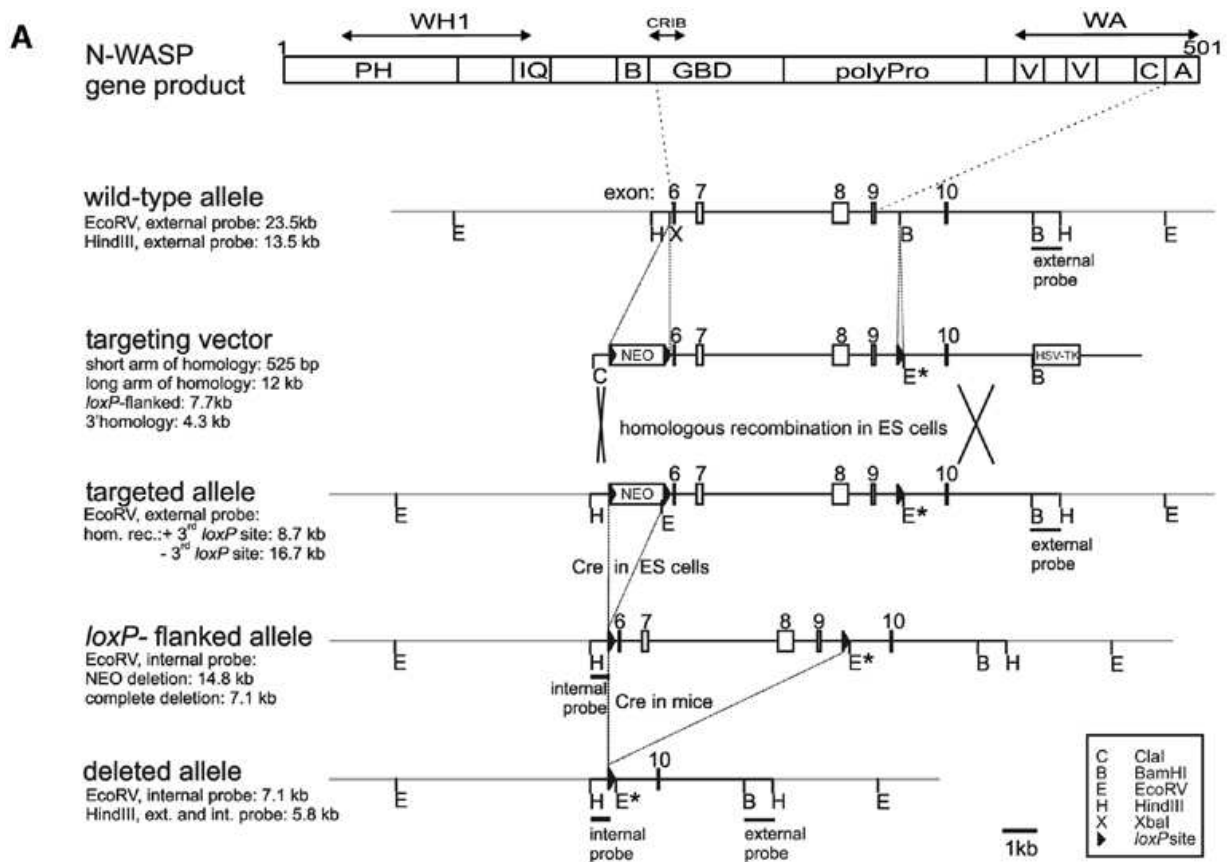
Actin filament assembly and turnover occurs in response to various extracellular stimuli and drives many cellular activities such as cell locomotion, cellular morphogenesis and phagocytosis. One prerequisite for actin reorganization is the *de novo* nucleation of actin filaments, which is also stimulated by intracellular pathogens that subvert the actin cytoskeleton and which have, therefore, been instrumental in identifying the essential

factors involved in this process (Frischknecht and Way, 2001). Members of the Wiskott–Aldrich Syndrome protein (WASP)/Scar family promote actin polymerization by stimulating the actin-nucleating activity of the Arp2/3 complex (reviewed in Takenawa and Miki, 2001). In contrast to Scar proteins, hematopoietic WASP and ubiquitously expressed N-WASP directly interact with the Rho-GTPase Cdc42 through their CRIB (Cdc42/Rac-interactive-binding) domain (Aspenstrom *et al.*, 1996; Rohatgi *et al.*, 1999). Cdc42 induces the formation of cell surface projections such as filopodia (Kozma *et al.*, 1995; Nobes and Hall, 1995), which is enhanced upon expression of N-WASP (Miki *et al.*, 1998). This suggests that filopodia formation results from Cdc42-based activation of N-WASP, which leads to Arp2/3 recruitment and subsequent actin filament assembly (Carlier *et al.*, 1999). In addition, WASP/N-WASP have been suggested to be involved in the formation of actin pseudopodia by enteropathogenic *Escherichia coli* (EPEC) (Kalman *et al.*, 1999), whereas intracellular *Shigella flexneri* and vaccinia virus have been shown to engage N-WASP for initiating actin polymerization at their surfaces in the host cell cytoplasm (Egile *et al.*, 1999; Frischknecht *et al.*, 1999).

In order to analyze N-WASP function in detail, we generated conditional knockout mice and blocked N-WASP expression during embryonic development. By establishing N-WASP-defective fibroblasts we were able to elucidate essential roles of N-WASP function at the cellular level.

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**Fig. 1.** Disruption of N-WASP leads to embryonic lethality. (A) Schematic representation of the N-WASP protein and the gene targeting strategy showing the genomic organization of the N-WASP locus in wild-type, targeted, floxed and deleted alleles. Also shown are the targeting vector and external and internal probes used for Southern blot analysis. Note the introduction of a new *EcoRV* restriction site upon gene targeting (marked *EcoRV*\*) used to distinguish the different alleles.

## RESULTS AND DISCUSSION

### N-WASP knockout leads to embryonic lethality

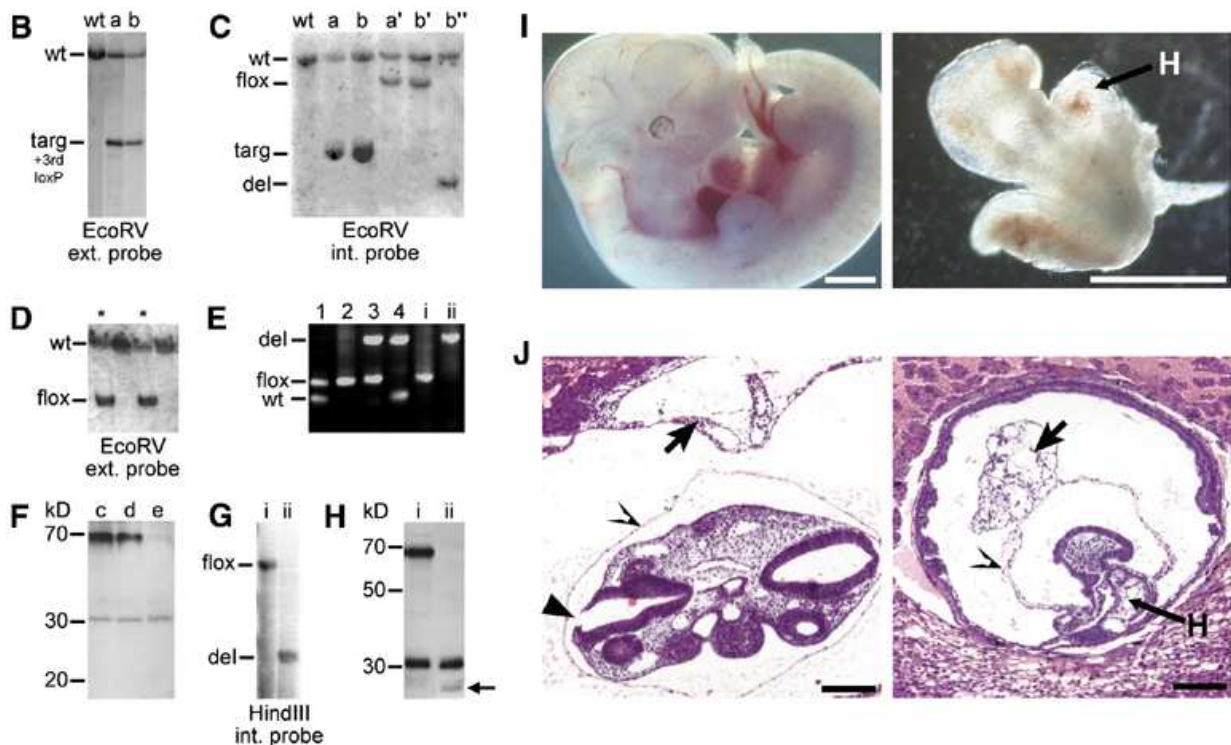
We generated conditionally gene targeted mice on a C57BL/6 background using the phage P1-derived *Cre/loxP* recombination system. To inactivate N-WASP we flanked the putative exons 6–9 with *loxP* sites by homologous recombination in embryonic stem (ES) cells (Figure 1A–C). After excision of the *neo*-selection marker in targeted ES cells by transient expression of Cre recombinase (Figure 1C), we derived germline targeted offspring carrying one *loxP*-flanked allele (N-WASP<sup>lox/+</sup>) (Figure 1D and E). N-WASP<sup>lox/lox</sup> mice behaved and bred normally showing no obvious alteration in N-WASP expression (Figure 1F). Excision of exons 6–9 by Cre eliminates N-WASP effector domains (Figure 1A) and splicing of exon 5 to the remaining exon 10 creates a premature stop codon (data not shown). After crossing with the transgenic mouse strain Cre159, which restricts Cre-mediated deletion to neurons in the forebrain (Minichiello *et al.*, 1999), expression of N-WASP protein was strongly reduced in extracts from forebrains of N-WASP<sup>lox/lox</sup> Cre159 mice as expected (Figure 1F), proving the successful targeting strategy. To assess the physiological consequences of N-WASP deficiency during embryonic development, we disrupted N-WASP in the

mouse germline by crossing with a deleter-Cre mouse strain (Schwenk *et al.*, 1995), leading to a constitutive N-WASP knockout (N-WASP<sup>del/del</sup>), which turned out to be embryonic lethal. Whereas heterozygous N-WASP<sup>del/wt</sup> mice appeared normal, no homozygous N-WASP<sup>del/del</sup> offspring were born. Thirty-nine implants were histologically analyzed in serial sections on embryonic day (E) 8.5, E9.0 and E9.5 and 79 embryos were dissected on days E9.5 to E13.5. Ten (25.6%) and 22 (27.8%) embryos, respectively, revealed developmental abnormalities and/or were partially resorbed. Upon genotyping of the dissected embryos by PCR, developmental abnormalities were found to correspond to the homozygous N-WASP<sup>del/del</sup> genotype. Thus, the abnormal embryos detected in the histological analysis shown in Figure 1J (see also Supplementary figure 1) most likely represent N-WASP<sup>del/del</sup> embryos. Figure 1I and J show a comparison of phenotypically normal and N-WASP<sup>del/del</sup> embryos demonstrating developmental retardation and reduced growth of N-WASP<sup>del/del</sup> embryos, which failed to undergo turning and showed abnormalities of intra- and extra-embryonal mesoderm differentiation (for further details see Supplementary data).

### Characterization of N-WASP<sup>del/del</sup> cell lines

Embryonic N-WASP<sup>lox/lox</sup> fibroblasts were immortalized, subcloned and various N-WASP<sup>del/del</sup> cell lines were selected after

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**Fig. 1. (continued)** (B and C) Southern analyses of precursor (wt), targeted (a and b), floxed (N-WASP<sup>flox/wt</sup>) (a', b'), and deleted (N-WASP<sup>del/del</sup>) (b'') ES clones. (D) Southern analysis of N-WASP<sup>flox/wt</sup> mice (marked with asterisks) derived after germline transmission. (E) PCR for genotypes of mice (lanes 1–4) and fibroblast cell lines (i and ii): wild-type, floxed and deleted alleles. Note the absence of the N-WASP<sup>flox</sup> allele in the N-WASP<sup>del/del</sup> cell line (ii). (F) Western blot analysis of N-WASP in extracts of hippocampi of wild-type (c), N-WASP<sup>flox/flox</sup> (d) and N-WASP<sup>flox/flox</sup> Cre159 mice (e) with polyclonal antibodies against the N-terminus of N-WASP. (G) Southern and (H) Western blot analyses of a representative N-WASP<sup>flox/flox</sup> fibroblast precursor cell line (i) and its corresponding N-WASP<sup>del/del</sup> cell line (ii). (H) Note the lack of N-WASP full-length expression in (ii). In both (F) and (H), the crossreaction at ~30 kDa was used as loading control. The small polypeptide of ~23 kDa detected in extracts of N-WASP<sup>del/del</sup> fibroblasts is indicated by an arrow. This product was not detected in forebrain extracts from Cre159 transgenic animals (F, lane e). (I) Comparison of genotyped embryos from intercrosses of N-WASP<sup>del/wt</sup> mice at E11.5. Note the reduced size and developmental retardation of the N-WASP<sup>del/del</sup> embryo (right) as compared with its wild-type sibling (left), H points to heart. Bars, 1 mm. (J) Histological analyses of N-WASP<sup>flox/flox</sup> embryo (left) and N-WASP<sup>del/wt</sup> embryo (right) at E9 from intercrosses of N-WASP<sup>flox/wt</sup> mice. The cranial parts of the wild-type embryo are well developed showing pharyngeal arches, a large brain cavity and an almost closed cranial neuropore in the hindbrain area (filled arrowhead). The heavily retarded embryo has not turned, but has formed a heart (marked H) and cranial mesenchyme, whereas the allantois (short arrow) shows widening of the intercellular spaces distally and does not contribute to the placenta as in wild-type. Open arrowheads point at amnion. Bars, 150  $\mu$ m.

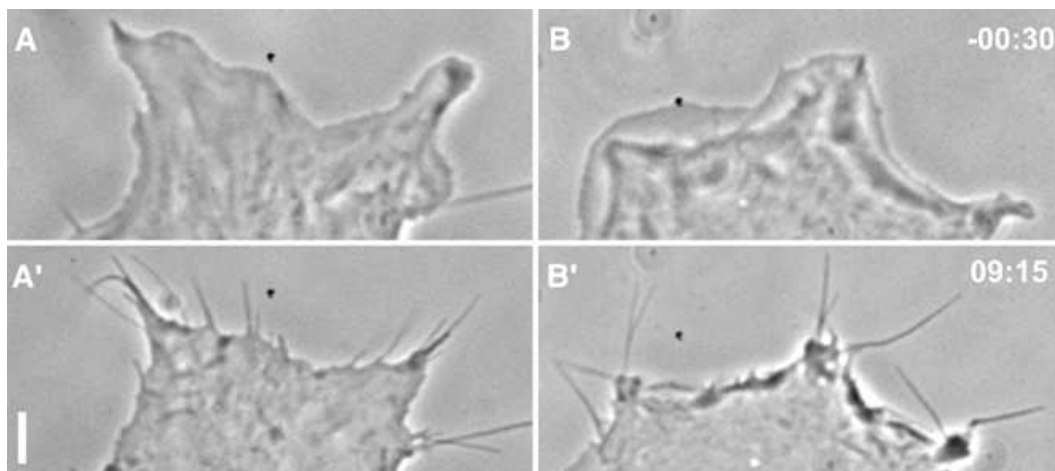
transient expression of Cre, thus enabling a direct comparison with their respective parental precursor lines. Figure 1G shows a Southern blot and Figure 1E shows PCR analysis of a representative N-WASP<sup>del/del</sup> cell line and its N-WASP<sup>flox/flox</sup> parental precursor line. Western blot analysis demonstrated lack of expression of full-length N-WASP in the N-WASP<sup>del/del</sup> cell line (Figure 1H). The weak labelling of a polypeptide with an estimated molecular weight of 23 kDa, which was recognized with two independent antibodies against the N-terminus of N-WASP, obviously corresponds to a truncated polypeptide (exons 1–5). However, we could not detect such a truncated gene product in extracts derived from conditionally N-WASP mutant mice (Figure 1F).

### Induction of filopodia by Cdc42 does not require N-WASP

Rho family GTPases have been recognized as key molecules in the regulation of actin-based cell motility (Hall, 1998), of which Cdc42 is thought to induce the formation of filopodia (Kozma et al., 1995; Nobes and Hall, 1995). Several effector proteins that

interact specifically with GTP-Cdc42 have been identified to date (Erickson and Cerione, 2001), but only N-WASP (Miki et al., 1998; Rohatgi et al., 1999) and the hematopoietic WASP (Symons et al., 1996) provide a direct link with actin assembly. The ubiquitously expressed N-WASP was therefore proposed to effect the formation of filopodia (Miki et al., 1998). Interestingly, none of our N-WASP-defective cell lines showed any apparent abnormalities in cell morphology compared with their respective precursor cell lines (not shown). We therefore sought to test whether the formation of filopodia induced by constitutively active Cdc42 is altered in N-WASP-defective cells. Nobes and Hall (1995) microinjected a mixture of constitutively active Cdc42, dominant-negative Rac (to suppress lamellipodia) and C3-transferase (to inhibit Rho), which caused the formation of filopodia in Swiss 3T3 fibroblasts. Microinjection of the same mixture into precursor (N-WASP<sup>flox/flox</sup>) and N-WASP-defective cells led to the rapid formation of numerous filopodia (Figure 2A and B, see also Supplementary video 1) in both cell types. Quantitation of filopodia ~30 min after microinjection revealed no difference between the average number of filopodia induced in N-WASP<sup>flox/flox</sup> ( $42 \pm 13$ ,  $n = 6$ ) and in N-WASP-defective cells





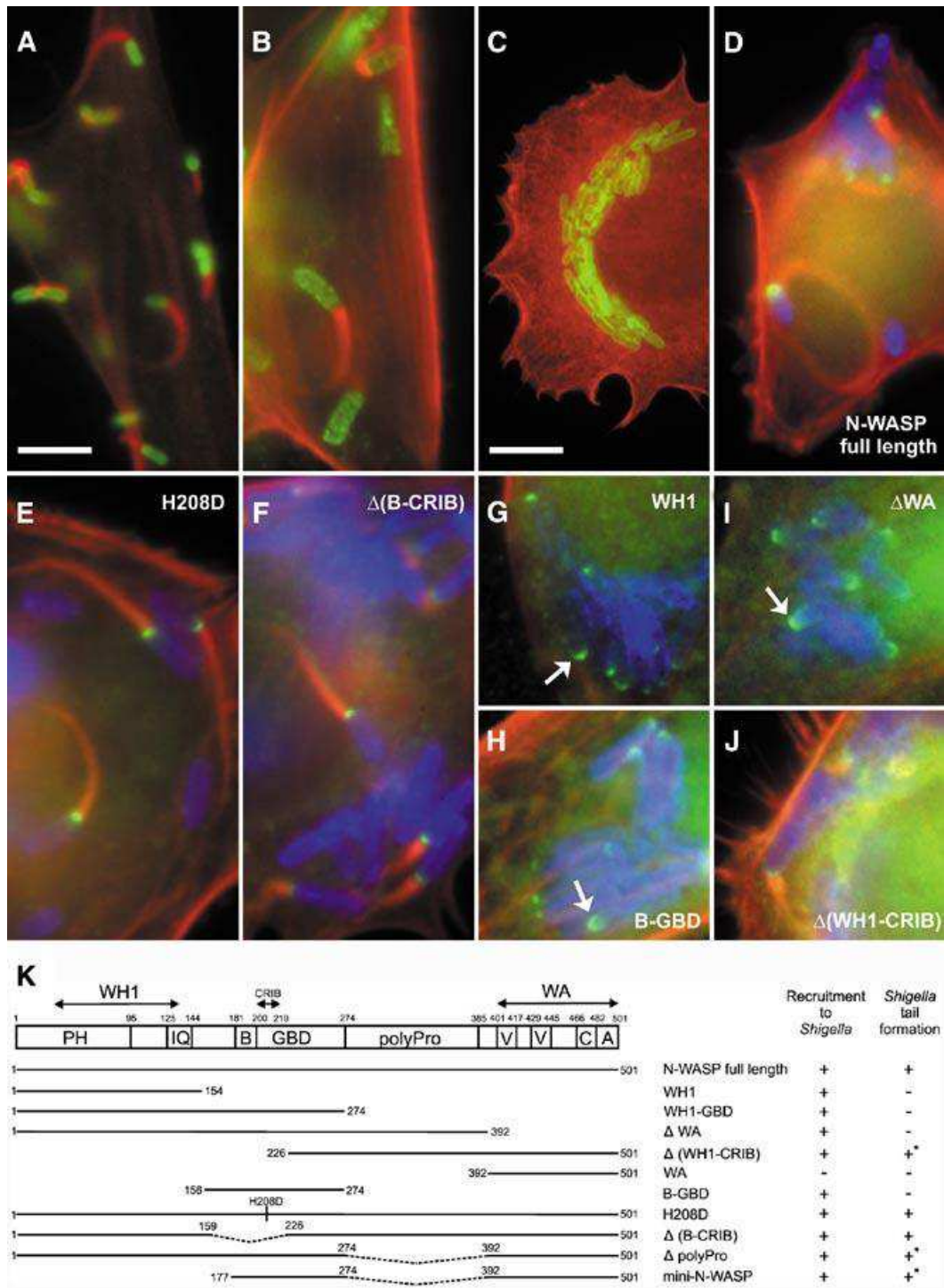
**Fig. 2.** N-WASP is not required for Cdc42-induced filopodia formation. Cells were microinjected with a mixture of L61Cdc42 (1.5mg/ml), N17Rac (0.35 mg/ml) and C3-transferase (0.1 mg/ml). Phase contrast images show a precursor (A, A') and a corresponding N-WASP-defective cell (B, B') before (A, B) and after (A', B') microinjection, which induced the formation of filopodia in both cell types. Approximate times before and after injections are given in minutes and seconds. Bar, 5  $\mu$ m.

( $48 \pm 16$ ,  $n = 9$ ). Thus, our data demonstrate for the first time that, at least in fibroblasts, N-WASP is not essential for the protrusion of filopodia induced by Cdc42.

### Infection of N-WASP-defective cells with intracellular bacterial pathogens

The bacterial pathogens *S. flexneri* and *Listeria monocytogenes* gain access to the host cytoplasm of infected cells where they induce the polymerization of actin filaments at their surfaces, which leads to the formation of comet-like actin tails, a process that provides the driving force for bacterial propulsion. In the case of *Shigella*, interaction of N-WASP with the bacterial surface protein IcsA/VirG is thought to unfold N-WASP leading to recruitment of the Arp2/3 complex and its activation, whereas the *Listeria* surface protein ActA recruits and activates the Arp2/3 complex directly (for references see Frischknecht and Way, 2001). In addition, the Rho-GTPase Cdc42 has been suggested to contribute to the actin-based motility of *Shigella* by initiating actin nucleation through formation of the IcsA/N-WASP/Arp2/3 complex (Suzuki *et al.*, 2000). When N-WASP-defective fibroblasts were infected with *S. flexneri* actin-tail formation was completely abolished (compare Figure 3C with B), whereas tail formation by *Listeria* was unaffected in these cells (Figure 3A). *Shigella*-induced actin tail formation could be fully restored upon ectopic expression of full-length murine N-WASP fused to GFP (Figure 3D) as well as by expression of N-WASP-H208D that is incapable of binding to GTP-Cdc42 (Figure 3E) (Miki *et al.*, 1998 and Supplementary figure 3) and a  $\Delta$ (B-CRIB) mutant (N-WASP- $\Delta$ aa160–225) (Figure 3F). In addition, time-lapse movies of *Shigella*-infected N-WASP-defective cells transfected with full-length N-WASP, N-WASP-H208D (see also Supplementary video 2) or with the  $\Delta$ (B-CRIB) mutant (not shown) revealed that the incapability of these mutant proteins to interact with Cdc42 (Supplementary figure 3) does not decrease *Shigella* motility. Since the H208D mutant of N-WASP was shown previously to be recruited to the *Shigella* surface (Suzuki *et al.*, 2000), it was suggested that IcsA and Cdc42 bind to different motifs in N-WASP

(Frischknecht and Way, 2001). Whereas our data confirm this hypothesis, they clearly demonstrate a recruitment mechanism independent of the CRIB domain. Furthermore, both the N-WASP construct B-GBD (GTPase binding domain) (aa 156–274, see Figure 3K) and the WH1 domain (aa 1–154) were recruited to the *Shigella* surface without inducing actin tails (Figure 3H and G, respectively), which was also observed when N-WASP- $\Delta$ WA was used as a control (Figure 3I). Previously, residues 148–273 of N-WASP were shown to target to the *Shigella* surface under experimental conditions where one can not exclude an interaction with endogenous N-WASP (Moreau *et al.*, 2000), since residues 209–274 were shown thereafter to interact intramolecularly with the C-terminal cofilin-homology-domain of N-WASP *in vitro* (Prehoda *et al.*, 2000). Our data refine the recruitment domain originally mapped by Moreau *et al.* (2000) to residues 226–273 within the GBD, but in addition suggest an alternative, WH1-domain-mediated recruitment mechanism, the nature and significance of which is currently unknown. Interestingly, constructs that lack the WH1 domain such as  $\Delta$ (WH1-CRIB) (Figure 3J) or mini-N-WASP (not shown, see summary in Figure 3K) were still capable of restoring actin tail formation, although average tail lengths appeared to be decreased as compared with full-length N-WASP. Whether this difference reflects a decrease in *Shigella* motility is currently under investigation. N-WASP lacking the polyproline region was still capable of restoring *Shigella*-induced actin tail formation, although again with less efficiency (not shown). This may be explained by the lack of direct interaction of the latter mutant with profilin I (Suet-sugu *et al.*, 1998), which has previously been shown to play a significant role in *Shigella* motility (Mimuro *et al.*, 2000). Together, in contrast to previous data (Suzuki *et al.*, 2000), we did not detect any apparent differences between *Shigella*-induced actin-tail formation upon rescue with full-length N-WASP as compared with H208D or  $\Delta$ (B-CRIB) mutants. We conclude that Cdc42 is not essential for the actin-driven motility of *Shigella*. Our conclusions corroborate both the observation that *Shigella*-induced actin tail formation is not impaired in infected cells treated with the clostridial toxin TcdB-10463, which blocks the



**Fig. 3.** N-WASP is essential for *Shigella*-induced actin tail formation. Precursor (B) and N-WASP-defective fibroblasts (A and C–J) were infected with *L. monocytogenes* (A) or *S. flexneri* (B–J). (A–C) show non-transfected cells, while (D–J) show cells expressing GFP-tagged N-WASP constructs as indicated. In all images, filamentous actin is shown in red. *Listeria* (A) and *Shigella* (B, C) are shown in green. In (D–J), *Shigella* and GFP fusion proteins are labeled blue and green, respectively. *Listeria* form actin tails in N-WASP-defective fibroblasts (A), while *Shigella*-induced actin tails are abolished in these cells (C), in contrast to the precursor cell line (B). Actin tail formation by *Shigella* is restored upon expression of full-length N-WASP (D), H208D, Δ(B-CRIB) and Δ(WH1-CRIB) mutants (E, F and J, respectively), but not after expression of N-WASP-WH1 (G), -B-GBD (H) or N-WASP-ΔWA (I), although the three latter constructs are recruited to the *Shigella* surface (indicated by arrows). Bar in (A) (5 μm) is valid for (A–J) except (C) (10 μm). (K) shows the domain structure of N-WASP and an overview of the GFP-tagged constructs used in this study. Recruitment to *Shigella* and reconstitution of actin tail formation are given. +\* marks low efficiency and/or shorter tails; PH, pleckstrin-homology domain; IQ, calmodulin-binding motif; B, basic domain; GBD, GTPase binding domain; polyPro, polyproline region; V, verprolin homology domain; C, cofilin homology domain and A, acidic domain.

GTPases of the Rho family including Cdc42 (Mounier *et al.*, 1999), and the fact that, in a reconstituted cell-free system, Cdc42 is not required for *Shigella*-motility (Loisel *et al.*, 1999).

### Enteropathogenic *E. coli* (EPEC)-induced formation of actin pedestals requires N-WASP

EPEC subvert the actin cytoskeleton by inducing the formation of characteristic pseudopodial structures (pedestals) with the bacteria residing at their tips (Goosney *et al.*, 2000). Over-expression of a WASP mutant lacking the C-terminal cofilin-homology region has been shown to interfere with pedestal formation in HeLa cells, an effect that was dependent on the CRIB domain of WASP (Kalman *et al.*, 1999). N-WASP, which localizes to the pedestal tips (Goosney *et al.*, 2001; this study), is therefore thought to be required for recruitment and activation of the Arp2/3 complex. It was further proposed that recruitment of WASP/N-WASP involves the Cdc42 homolog Chp (Aronheim *et al.*, 1998; Kalman *et al.*, 1999). When N-WASP-defective fibroblasts were infected with EPEC we found that, although the bacteria readily attached to the cell surface (Figure 4C), pedestal formation was completely blocked (Figure 4D) in contrast to infected precursor cells (Figure 4A and B). Pedestal formation could be fully restored in N-WASP-defective cells by expression of full-length N-WASP (Figure 4E), N-WASP-H208D (Figure 4F) or the  $\Delta$ (B-CRIB) mutant (Figure 4G), but not by expression of N-WASP mutants truncated at the N-terminus or lacking the WA-effector-domain (summarized in Figure 4K). Furthermore, the WH1-GBD construct (aa 1–274) clearly targeted to the host cell surface (Figure 4I) at sites of EPEC attachment (Figure 4H) without inducing formation of actin pedestals (Figure 4J). Thus, the recruitment domain of N-WASP is restricted to its N-terminal half. A construct lacking the polyproline region ( $\Delta$ polyproline, see Figures 3K and 4K) also restored pedestal formation, suggesting that a direct interaction of N-WASP with profilin I or SH3-domain containing adaptor proteins such as Grb2 (Carlier *et al.*, 2000), shown previously to be recruited to EPEC pedestals (Goosney *et al.*, 2001), is not essential for this process. Mini-N-WASP, an N-terminally truncated version of the latter construct (see Figure 3K and 4K), which corresponds to a mutant that could still be induced by PIP<sub>2</sub> (phosphatidylinositol-4-phosphate) and Cdc42 to activate the Arp2/3 complex *in vitro* (Prehoda *et al.*, 2000), did not restore pedestal formation. Therefore we are currently investigating the role of the WH1 domain in N-WASP targeting to the attachment site of EPEC, although so far it has been difficult to detect a recruitment of the WH1 domain alone (not shown). In summary, in contrast to previous results (Kalman *et al.*, 1999), this study clearly shows that recruitment of N-WASP to attachment sites of EPEC at the host cell membrane is not mediated by the CRIB-domain.

### Speculation

N-WASP is a highly potent activator of the Arp2/3 complex *in vitro* and harbors key regulatory functions for cellular actin assembly. Our study proves the essential role of N-WASP, since disruption of the gene leads to embryonic lethality. In addition, two out of three bacterial pathogens investigated in this study have selected N-WASP as the target molecule to subvert the host cell actin polymerization machinery. This may be due to the

presence of an additional verprolin homology region in the C-terminus of N-WASP, which has been correlated with the higher potency of N-WASP to activate the Arp2/3 complex compared with the highly related Scar/WASP family members (Yamaguchi *et al.*, 2000). However, the precise *in vivo* functions of N-WASP remain to be determined, since it is not clear whether the early embryonic lethality is due solely to disturbances in actin dynamics or may be caused by additional, so far unknown, activities of N-WASP. Nevertheless, the availability of conditionally N-WASP-targeted mice not only enables the generation of specific cell lines for analyzing its role in actin-based motility as described above, but is also a prerequisite for elucidating the physiological functions of N-WASP *in vivo* by alteration of its temporal and tissue-specific expression.

## METHODS

**Transgenic animals.** Conditional gene targeting and generation of transgenic animals were performed according to standard protocols and as described in the Supplementary material.

**Cell culture and media.** Primary embryonic fibroblasts from N-WASP<sup>fllox/fllox</sup> mice were prepared on E14 cells and immortalized with retroviruses causing expression of a temperature-sensitive simian virus 40 large-T-antigen (Jat and Sharp, 1989). Fibroblasts were grown at 32°C in DMEM with glutamine and 10% FCS (Life Technologies). Transfections were performed with FuGENE6 (Roche).

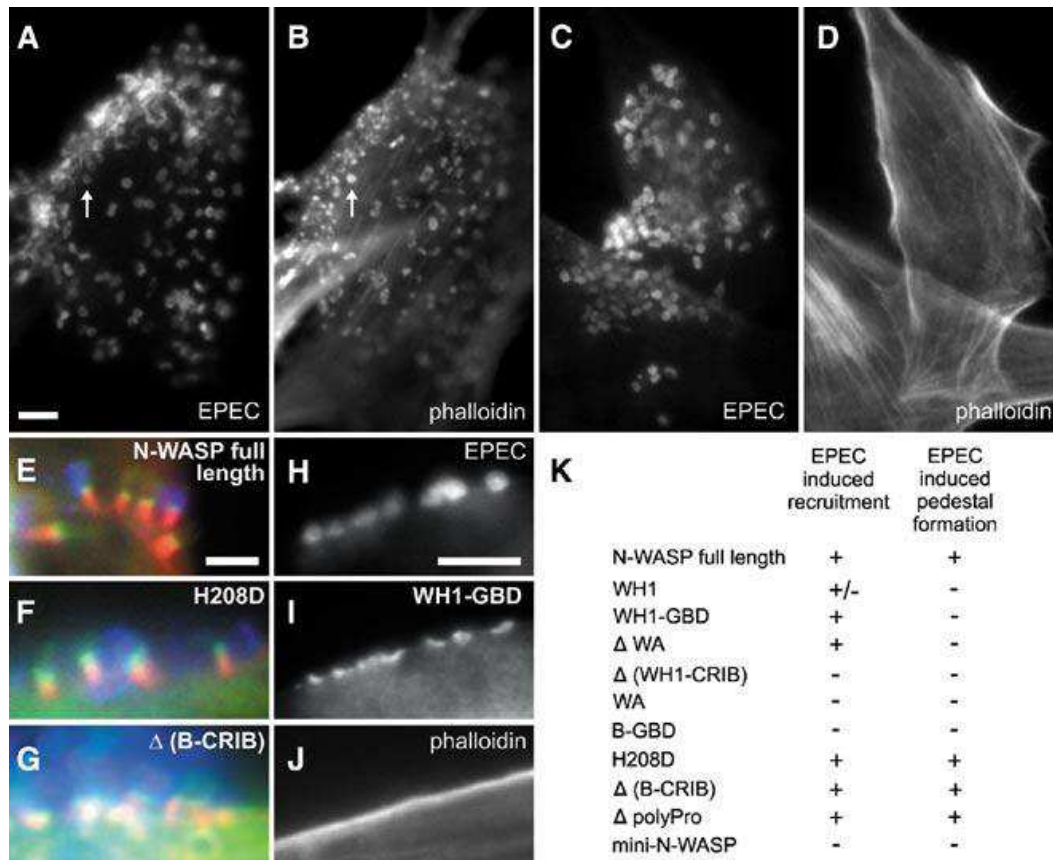
**N-WASP expression constructs.** We isolated the murine N-WASP cDNA by performing independent RT-PCR on RNA from brains of C57BL/6 mice using primers 5'-AGAGTGGGACCCGAGTGCTC-3' and 5'-ACAGATCAATAGACAGGAAG-3'. For GFP-tagged N-WASP expression constructs, murine N-WASP cDNA regions were cloned into pEGFP-C1 or -C2 vectors (Clontech), sequenced and expression was verified by Western blotting (see Supplementary figure 2).

**Bacterial strains and infections.** We used EPEC strain 2348/69, M90T wild-type invasive strain of *S. flexneri* serotype 5 and *L. monocytogenes* EGD serotype 1/2a. For EPEC infections, overnight cultures were diluted 1:50 into DMEM and grown for 3 h at 37°C. After diluting 1:100, bacteria were centrifuged onto fibroblasts at 700 g for 5 min and incubated for 4–5 h with hourly medium changes. Prior to infections with *S. flexneri*, bacteria grown overnight in trypticase soya bouillon (TSB) were diluted 1:100 into DMEM and centrifuged onto cells. One and a half hours after infection, DMEM was exchanged for DMEM with 10% FCS and 50 µg/ml gentamicin (Sigma-Aldrich) and incubated for 2 h. *Listeria* infections were performed as described (Carl *et al.*, 1999).

**Immunofluorescence.** Indirect immunofluorescence was performed essentially as described (Herzog *et al.*, 1994). We used polyclonal antibodies against *S. flexneri*, monoclonal antibodies against EspE, both provided by T. Chakraborty (Giessen, Germany), polyclonal antibodies against *L. monocytogenes* (Domann *et al.*, 1992). Secondary reagents were Alexa-Fluor 488-conjugated goat anti-rabbit (Molecular Probes) and AMCA-conjugated goat anti-mouse antibodies (Dianova), which were mixed with Alexa 594-phalloidin (Molecular Probes) to stain filamentous actin.

**Microinjection, video microscopy and image processing.** Recombinant L61-Cdc42, N17Rac and C3-transferase were





**Fig. 4.** EPEC pedestal formation depends on N-WASP. Infection of precursor (A, B) and N-WASP-defective cells (C–J) with EPEC. EPEC are shown in (A, C and H) and in blue in (E–G). F-actin is shown in (B, D and J) and in red in (E–G). GFP constructs are shown in (I) and in green in (E–G). EPEC (A, C)-induced formation of actin pedestals in precursor (B), but not in N-WASP-defective cells (D). Pedestal formation in N-WASP-defective cells was restored upon expression of GFP-tagged full-length N-WASP (E), N-WASP-H208D (F) and N-WASP-Δ(B-CRIB) (G), while N-WASP-WH1-GBD was recruited (I) by EPEC (H) without induction of actin pedestals (J). Bars (5 μm) in (A, E and H) are valid for (A–D), (E–G) and (H–J), respectively. For description of GFP fusion constructs see Figure 3K. (K) The recruitment to EPEC attachment sites and reconstitution of pedestal formation by the respective mutants are indicated.

purified and injected in a mixture essentially as described previously (Nobes and Hall, 1995). Time-lapse movies and fluorescence images were acquired and processed as described (Rottner et al., 1999).

**Accession numbers.** The murine N-WASP cDNA sequence has been submitted to the DDBJ/EMBL/GenBank Nucleotide Sequence Database under accession No. AJ318416.

**Supplementary data.** Supplementary data are available at *EMBO reports* Online.

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